Code No. 27419

Mouse/Rat sAPPα (highly sensitive) Assay Kit - IBL

INTRODUCTION

Alzheimer's disease (AD) was first reported by A. Alzheimer, a German neuropathologist in 1907 and is considered as a major factor of dementia. It is known that Amyloid β (Aβ; which is major constituent of senile plaque) is cleaved from Amyloid Precursor Protein (APP; which exists in three main isoforms, APP695, APP751, and APP770) by β-secretase and subsequent γ-secretase (ref. 1).
The production of soluble APPβ (sAPPβ) by β-secretase cleavage corresponds to Aβ production accordingly, so it is desired to measure sAPPβ in parallel with Aβ.
On the other hand, it is considered that in the metabolic pathway of APP, APP is first cleaved by α-secretase rather than β-secretase normally to produce soluble APPα (sAPPα) and subsequently P3 is cleaved from the remaining C-terminal fragment by γ-secretase. In recent research, there are several attempts to apply the inhibitor of β-secretase and the activation of α-secretase for AD treatment.
As stated above, it is useful for research of AD to measure sAPPα as well as sAPPβ.
This kit can measure Mouse/Rat sAPPα.
Note: Please pay attention in sample selection since this kit also measures full-length APP.

PRINCIPLE

This kit is a solid phase sandwich ELISA using 2 kinds of highly specific antibodies. Tetra Methyl Benzidine (TMB) is used as a coloring agent (Chromogen). The strength of coloring is proportional to the quantities of Mouse/Rat sAPPα.

MEASUREMENT RANGE

4.1 - 260 pg/mL

INTENDED USE

For research use only, not for use in diagnostic procedures.

This IBL's assay kit is capable for the quantitative determination Mouse/Rat sAPPα in serum, EDTA plasma and cell culture supernatant. Guideline of dilution for serum/plasma samples of normal mouse/rat is around 50-fold.
Note: Please pay attention in measuring of the other samples since this kit also detects full-length APP molecules.

KIT COMPONENT

1	Precoated plate	: Anti-Mouse APP(599) Rabbit IgG Affinity Purity	96Well x 1
2	Labeled antibody Conc.	:	
	(30X) HRP conjugated	Anti- Mouse N-APP Rabbit IgG Fab' Affinity Purify	0.4mL x 1
3	Standard	: Recombinant mouse sAPPα	0.5mL x 2
4	EIA buffer	: 1% BSA, 0.05% Tween20 in PBS	30mL x 1
5	Solution for Labeled antibody	: 1% BSA, 0.05% Tween20 in PBS	12mL x 1
6	Chromogen	: TMB solution	15mL x 1
7	Stop solution	: 1N H_2SO_4	12mL x 1
8	Wash buffer Conc.	: (40X) Phosphate buffer	50mL x 1

OPERATION MANUAL

1. Materials needed but not supplied

- Plate reader (450nm)
- Micropipette and tip
- Graduated cylinder and beaker
- Deionized water
- Refrigerator (as 4°C)
- Graph paper (log/log)
- Paper towel
- Tube for dilution of Standard
- Washing bottle for precoated plate
- Disposable test tube for "2, Labeled antibody Conc." and "6, Chromogen"

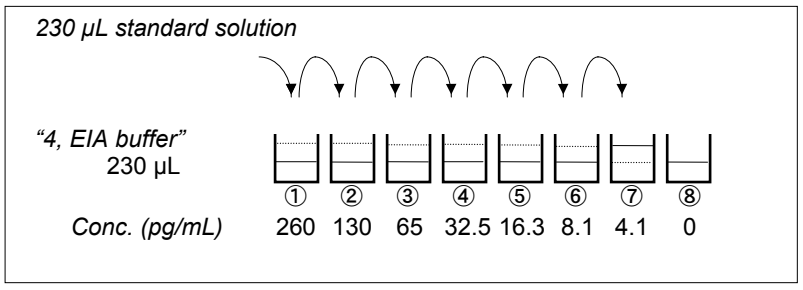
2. Preparation

1) Preparation of wash buffer
"8, Wash buffer Conc." is a concentrated (40X) buffer. Adjust the temperature of "8, Washing buffer Conc." to room temperature and then, mix it gently and completely before use. Dilute 50 mL of "8, Wash buffer Conc." with 1,950 mL of deionized water and mix it. This is the wash buffer for use. This prepared wash buffer shall be stored in refrigerator and used within 2 weeks after dilution.
2) Preparation of Labeled antibody
"2, Labeled antibody Conc." is a concentrated (30X). Dilute "2, Labeled antibody Conc." with "5, Solution for Labeled antibody" in 30 times according to required quantity into a disposable test tube. Use this resulting solution as Labeled antibody.
Example)
In case you use one strip (8 well), the required quantity of Labeled antibody is 800 µL. (Dilute 30 µL of "2, Labeled antibody Conc." with 870 µL of "5, Solution for Labeled antibody" and mix it. And use the resulting solution by 100 µL in each well.)
This operation should be done just before applying labeled antibody.
The remaining "2, Labeled antibody Conc." should be stored at 4°C in firmly sealed vial.
3) Preparation of Standard
Put just 0.5 mL of deionized water into the vial of "3, Standard" and mix it gently and completely. This solution is 520 pg/mL Mouse/Rat sAPPα standard.
4) Dilution of Standard
Prepare 8 tubes for dilution of "3, Standard". Put 230 µL each of "4, EIA buffer" into the tube.
Specify the following concentration of each tube."

Tube-1	260 pg/mL
Tube-2	130 pg/mL
Tube-3	65 pg/mL
Tube-4	32.5 pg/mL
Tube-5	16.3 pg/mL
Tube-6	8.1 pg/mL
Tube-7	4.1 pg/mL
Tube-8	0 pg/mL (Test Sample Blank)

Put 230 µL of Standard solution into tube-1 and mix it gently. Then, put 230 µL of tube-1 mixture into tube-2. Dilute two times standard solution in series to set up 7 points of diluted standard between 260 pg/mL and 4.1 pg/mL. Tube-8 is the test sample blank as 0 pg/mL.

See following picture.



5) Dilution of test sample
Test samples should be diluted with "4, EIA buffer" suitably.
Guideline of dilution for serum/plasma samples of normal mouse/rat is around 50-fold.

3. Measurement procedure

All reagents shall be brought to room temperature approximately 30 minutes before use. Then mix it gently and completely before use. Make sure of no change in quality of the reagents. Standard curve shall be prepared simultaneously with the measurement of test samples.

Reagents	Test Sample	Standard	Test Sample Blank	Reagent Blank
	Test sample 100 µL	Diluted standard (Tube 1-7) 100 µL	EIA buffer (Tube-8) 100 µL	EIA buffer 100 µL
Incubation overnight at 4 °C with plate lid				
Washing 4 times				
Labeled Antibody	100 µL	100 µL	100 µL	-
Incubation for 60 minutes at 4°C with plate lid				
Washing 5 times				
Chromogen	100 µL	100 µL	100 µL	100 µL
Incubation for 30 minutes at room temperature (shielded)				
Stop solution	100 µL	100 µL	100 µL	100 µL
Read the plate at 450nm against a Reagent Blank within 30 minutes after addition of Stop solution.				

1) Determine wells for reagent blank. Put 100 µL each of "4, EIA buffer" into the wells.
2) Determine wells for test sample blank, test sample and diluted standard.
Then, put 100 µL each of test sample blank (tube-8), test sample and dilutions of standard (tube-1-7) into the appropriate wells.
3) Incubate the precoated plate overnight at 4°C after covering it with plate lid.
4) Wash each well of the precoated plate 4 times with wash buffer using a washing bottle or a plate washer in following way.
After shaking off (or aspiration of) the solution in wells, fill each well with wash buffer and shake off the wash buffer completely from the precoated plate. This procedure must be repeated 4 times. Then, drain the precoated plate completely on paper towel.
Please refer to 5) and 6) in SPECIAL ATTENION below, and be careful not to miss a well.
5) Pipette 100 µL of labeled antibody solution into the wells of test samples, diluted standard and test sample blank.
6) Incubate the precoated plate for 60 minutes at 4°C after covering it with plate lid.
7) Wash the precoated plate 5 times in the same manner as 4).
In case of using a plate washer, we recommend manually washing in the manner mentioned above at least the last one time.
8) Take the required quantity of "6, Chromogen" and put it into a disposable test tube. Then, pipette 100 µL from the test tube into every well. Please do not return the rest of used chromogen in the test tube into "6, Chromogen" bottle in order to avoid contamination.
9) Incubate the precoated plate for 30 minutes at room temperature in the dark. The solution of Chromogen will turn blue.
10) Add 100 µL of "7, Stop solution" to all wells. Mix the solution by tapping the side of precoated plate. The solution will turn yellow by addition of "7, Stop solution".
11) Remove any dirt or drop of water on the bottom of the precoated plate and confirm there is no bubble on the surface of the solution. Then, run the plate reader and conduct measurement at 450 nm against a reagent blank.
The measurement shall be done within 30 minutes after addition of "7, Stop solution".

SPECIAL ATTENTION

1) Test samples should be measured soon after collection. For the storage of test samples, store them frozen and do not repeat freeze/thaw cycles. Thaw the test samples at a low temperature and mix them completely before measurement.
2) Test samples should be diluted with "4, EIA buffer", suitably.
3) Duplicate measurement of test samples and standard is recommended.
4) Use test samples in neutral pH range. The contaminations of organic solvent may affect the measurement.
5) Use only wash buffer in this kit for washing the precoated plate.
Insufficient washing may lead to the failure in measurement.

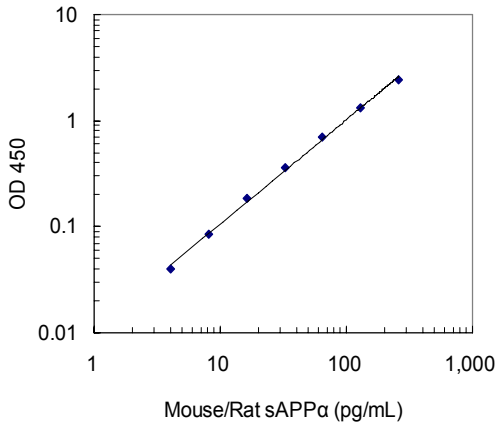
6) Remove the wash buffer completely by tapping the precoated plate on paper towel. Do not wipe wells with paper towel.
7) "6, Chromogen" should be stored in the dark due to its sensitivity against light. Avoid contact of Chromogen with metals.
8) Measurement should be done within 30 minutes after addition of "7, Stop solution".

CALCULATION OF TEST RESULT

Subtract the absorbance of test sample blank from all data, including standards and unknown samples before plotting. Plot the subtracted absorbance of the standards against the standard concentration on log-log graph paper. Draw the best smooth curve through these points to construct the standard curve. Read the concentration for unknown samples from the standard curve.

Example of standard curve

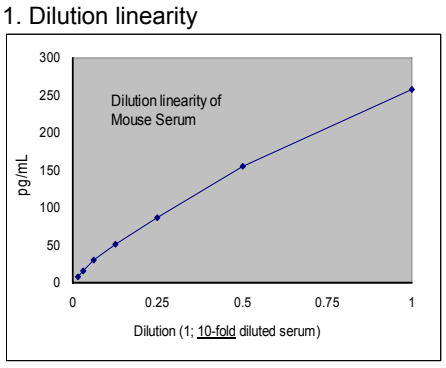
Conc. (pg/mL)	Absorbance (450nm)
260	2.421
130	1.328
65	0.721
32.5	0.385
16.3	0.205
8.1	0.107
4.1	0.061
0 (Test Sample Blank)	0.021

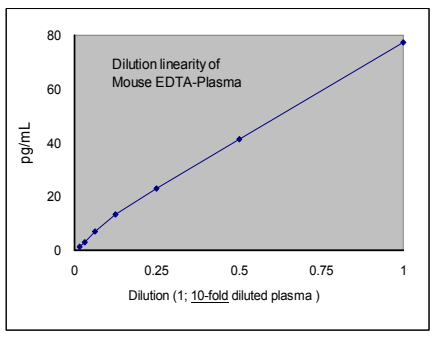


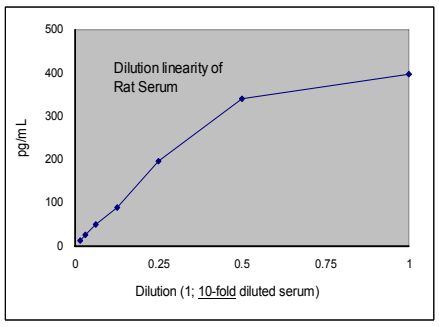
∗ The typical standard curve is shown above. This curve can not be used to derive test results. Please run a standard curve for each assay.

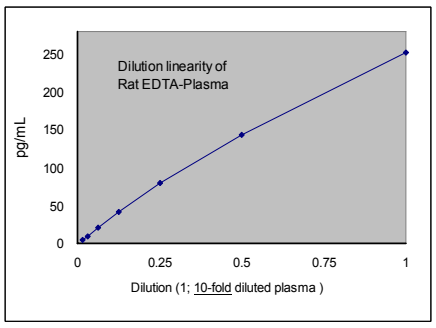
PERFORMANCE CHARACTERISTICS

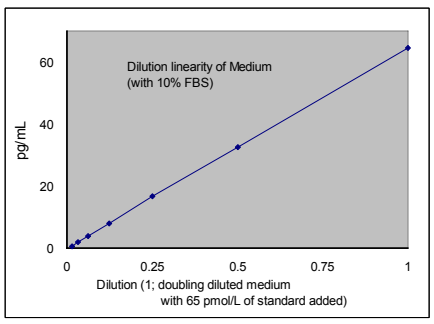
1. Dilution linearity











2. Added Recovery Assay

Specimen	Additive Amount (pg/mL)	Theoretical Value (pg/mL)	Measured Value (pg/mL)	%
Mouse Serum (BALB/c) (x50)	65.0	143.0	104.0	72.7
	16.3	94.2	80.6	85.6
	4.1	82.0	73.5	89.6
Mouse Plasma (EDTA) (BALB/c) (x50)	32.5	63.9	60.1	94.1
	8.1	39.5	40.6	102.8
	2.0	33.4	33.8	101.2
Rat Serum (x50)	65.0	245.9	222.4	90.4
	32.5	213.4	197.4	92.5
	4.1	184.9	184.9	100.0
Rat Plasma (EDTA) (x50)	32.5	92.0	91.9	99.9
	16.3	75.7	78.1	103.2
	4.1	63.5	62.9	99.1
Medium with 10% FBS (x2)	65.0	65.1	63.0	96.8
	16.3	16.3	15.8	96.9
	2.0	2.1	1.9	90.5

3. Intra - Assay

Mean Value (pg/mL)	SD (pg/mL)	CV (%)	n
154.0	3.1	2.0	24
65.9	1.1	1.7	24
27.3	0.6	2.2	24

4. Inter - Assay

Mean Value (pg/mL)	SD (pg/mL)	CV (%)	n
168.4	9.9	5.9	4
73.0	4.8	6.6	4
29.4	1.4	4.8	4

5. Specificity

Substance	Cross-Reactivity
Mouse/Rat sAPPα	100 %
Mouse/Rat sAPPβ	0.15 %

6. Sensitivity

0.6 pg/mL
The sensitivity for this kit was determined using the guidelines under the National Committee for Clinical Laboratory Standards (NCCLS) Evaluation Protocols. (National Committee for Clinical Laboratory Standards Evaluation Protocols, SC1, (1989) Villanova, PA: NCCLS.)

PRECAUTION FOR INTENDED USE AND/OR HANDLING

1. All reagents should be stored at 2 - 8°C. All reagents shall be brought to room temperature approximately 30 minutes before use.
2. "3, Standard" is lyophilized products. Be careful to open this vial.
3. "7, Stop solution" is a strong acid substance. Therefore, be careful not to have your skin and clothes contact "7, Stop solution" and pay attention to the disposal of "7, Stop solution".
4. Dispose used materials after rinsing them with large quantity of water.
5. Precipitation may occur in "2, Labeled antibody Conc.", "4, EIA buffer" or "8, Wash buffer Conc.", however, there is no problem in the performance.
6. Wash hands after handling reagents.
7. Do not mix the reagents with the reagents from a different lot or kit.
8. Do not use expired reagents.
9. This kit is for research purpose only. Do not use for clinical diagnosis.

STORAGE AND THE TERM OF VALIDITY

Storage Condition : 2 - 8°C
The expiry date is specified on outer box.

REFERENCE

1. Selkoe DJ. Normal and abnormal biology of the beta-amyloid precursor protein.Annu Rev Neurosci. 1994;17:489-517.
2. Citron M, Oltersdorf T, Haass C, McConlogue L, Hung AY, Seubert P, Vigo-Pelfrey C, Lieberburg I, Selkoe DJ. Mutation of the beta-amyloid precursor protein in familial Alzheimer's disease increases beta-protein production. Nature. 1992 Dec 17;360(6405):672-4.
3. Hsiao K, Chapman P, Nilsen S, Eckman C, Harigaya Y, Younkin S, Yang F, Cole G. Correlative memory deficits, Abeta elevation, and amyloid plaques in transgenic mice. Science. 1996 Oct 4;274(5284):99-102.

Version 1.

Made in Japan.